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## COMMENTARY

## Drugs and trafficking of ion channels: a new pro-arrhythmic threat on the horizon?

MAG van der Heyden, ME Smits and MA Vos

Division of Heart & Lungs, Department of Medical Physiology, UMC Utrecht, The Netherlands

Tuning of functional expression levels of the ion channels that make up the cardiac action potential (AP) is crucial for preserving correct AP duration (APD) and QTc times. Many compounds inhibit human ether-à-go-go related gene (hERG)-mediated delayed rectifier currents and thus prolong cardiac repolarization that may cause life-threatening arrhythmias like Torsades de Pointes. An increasing number of drugs are found to inhibit hERG function by a dual mechanism of short-term channel block and long-term trafficking defects that operate over different time and concentration scales. In safety screens at present used by pharmaceutical companies, the short-term effect of channel block is covered. In contrast, specific screening for long-term trafficking defects is not common, with the consequent risk of drugs that disturb trafficking entering the market. Whether that poses another pro-arrhythmic threat for the patients treated has to be determined, but is not unlikely. British Journal of Pharmacology (2008) 153, 406-409; doi:10.1038/sj.bjp.0707618; published online 3 December 2007

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Abbreviations: AP, action potential; APD, action potential duration; hERG, human ether-à-go-go related gene;  $I_{Kr}$ , rapid delayed rectifier current carried by hERG; LQTS, long QT syndrome; QT, depolarizing-repolarizing time interval on ECG

The electrocardiogram (ECG) is the result of voltage differences generated by tens of millions of cellular action potentials (AP). In mammalian cardiac ventricular myocytes, the AP is itself based on many inward and outward currents. The principal repolarization currents are the rapidly  $(I_{Kr})$  and slowly  $(I_{Ks})$  activating components of the delayed rectifier K<sup>+</sup> current. The pore-forming subunits of channels responsible for  $I_{Kr}$  are encoded by the human ether-à-go-go related gene (hERG or KCNH2) and for  $I_{Ks}$  by the KCNQ1 gene, while modulating proteins encoded by the KCNE family are also implicated. Mutations in these genes were initially linked to repolarization disorders (the congenital long depolarizingrepolarizing time interval on ECG (QT) syndrome, LQTS), to the occurrence of torsades de pointes arrhythmias and to sudden cardiac death, often when the repolarization reserve is maximally challenged (Roden et al., 2002).

The starting paradigm was that these mutations were causing malfunctioning of the specific repolarization currents ('loss of  $I_{Kr}$  function'), thereby decreasing the total current available and increasing the time necessary for repolarization to complete. More recently, it has been shown that many if not most of the LQTS2 mutations are caused by

trafficking defects instead of malfunctioning of the current (Anderson et al., 2006). By decreasing the number of proteins that reach the sarcolemma, the number of ion channel proteins is diminished ('reduced hERG density') also leading to less total current and hence to problems in controlling

At the other end of the spectrum of the LQTS are drugs that block specific ion channels, reducing the current and prolonging the cellular AP duration (APD) visualized on the ECG as an increase in QT time (Fitzgerald and Ackerman, 2005). These drugs have also been implicated in torsades de pointes arrhythmias and sudden cardiac death (Thomsen et al., 2006). The list of drugs that are pro-arrhythmic is alarmingly long and consists mostly of  $I_{Kr}$  blockers ('block of function') that have common drug binding sites within the protein structure (Stansfeld et al., 2007). More recently, attention has shifted to drugs that disrupt hERG protein trafficking to the cell surface membrane (for example Cordes et al., 2005; Kuryshev et al., 2005; Rajamani et al., 2006; Sun et al., 2006; Guo et al., 2007; Wang et al., 2007). An extensive screen performed by Wible et al. (2005) showed that approximately 40% of acute hERG blockers also affect trafficking leading to the 'reduced density' hypothesis to explain their pro-arrhythmic mechanism (Eckhardt et al., 2005; Hancox and Mitcheson, 2006). Of note, there are drugs that share both properties and in case of the antifungal drug ketoconazole, the contribution is equal, independent and additive as described in this issue of the British Journal of Pharmacology by Takemasa et al. (2007).

Correspondence: Dr MAG van der Heyden, Department of Medical Physiology, DH&L, UMC Utrecht, Yalelaan 50, Utrecht 3584 CM, The Netherlands.

E-mail: m.a.g.vanderheyden@umcutrecht.nl

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One of the characteristics that may distinguish between the two mechanisms of action is the time at which the drug effect becomes evident. In case of 'block of function', the block appears immediately (minutes), while in the 'reduced density' hypothesis, this action may take from hours to days to occur. In the Takemasa study, the 'block of function' effect is maximal within 5 min following drug application. The 'reduced density' effect becomes apparent 8 h after treatment and mature hERG expression levels continue to decrease for up to 48 h. Another, clinically important, observation indicated that upon washout of ketoconazole, the 'block of function' was resolved within minutes, while the 'reduced density' persisted for up to 48 h.

A second characteristic difference between the two effects is the underlying molecular mechanism. As shown by Takemasa *et al.* (2008) and Ridley *et al.* (2006), 'block of function' depends on two aromatic amino-acid residues, Y652 and F656, residing in the transmembrane region S6 of the hERG channel, while 'reduced density' is independent of them. Ketoconazole affected hERG maturation and therefore impaired trafficking from ER to Golgi (Takemasa *et al.*, 2008) and these authors strengthened their findings by applying fluorescence-activated cell sorting and electrophysiology to demonstrate that fewer hERG channels reach the plasma membrane. Importantly, they declined to rely on suggestive

but inconclusive confocal microscopy, which inherently suffers from insufficient spatial resolution to distinguish between localization on or just below the plasma membrane. Data from different fields indicate the involvement of specific amino-acid domains and interacting accessory proteins for effective trafficking of cardiac ion channels towards the sarcolemma (Steele *et al.*, 2007). As many of the congenital forms of LQT2 involve trafficking defects, analysis of the function of these mutated residues (Walker *et al.*, 2007) is a first step to elucidate the mode of action by which drugs affect trafficking of cardiac ion channels.

This difference in mechanism between 'block of function' and 'reduced density' poses a fundamental question. Can long-term trafficking defects occur without short-term blockade? In the current drug testing assays, agents are considered as non-safe when they induce  $I_{\rm Kr}$  block acutely and these compounds are often not pursued further. This may be an enormous waste, since  $I_{\rm Kr}$  block does not necessarily result in APD/QT-lengthening or life-threatening arrhythmias *in vivo*. Nevertheless, this approach will prevent potentially unsafe drugs from entering the market. On the other hand, drugs that display only long-term effects might pass the short-term safety screens. Some of these drugs may be eliminated as a result of long-term *in vivo* screening, but only when they cause QT prolongation,

**Table 1** Drugs evaluated for short and long term effects on  $I_{Kr}$ 

Drug	I <sub>kr</sub> channel block	hERG trafficking inhibition	Ratio	Other currents <sup>a</sup>	Trafficking others <sup>b</sup>	QT-APD increase	Reference
Arsenic trioxide	+	++	> trafficking	$I_{CaL} \uparrow$ $I_{Ks} \downarrow \longleftrightarrow$ $I_{Kur} \uparrow$ $I_{K-ATP} \uparrow$ $I_{to} \longleftrightarrow$ $I_{K1} \longleftrightarrow$ $I_{Na} \longleftrightarrow$	Kv1.5↑	Yes	Drolet <i>et al.</i> (2004) Ficker <i>et al.</i> (2004)
Celastrol	+	+ +	> trafficking	$I_{K1}\downarrow$ $I_{Kv2.1}\leftrightarrow$	Kir2.1↓	Yes	Sun et al. (2006)
Digitoxin/ouabain/ digoxin	_	+	> trafficking	$I_{CaL} \leftrightarrow$ $I_{Ks} \leftrightarrow$ $I_{Kur} \leftrightarrow$	Kv1.5 ↔	Yes	Wang <i>et al</i> . (2007)
Dofetilide	+	Not reported		I <sub>K1</sub> ↔ Specific I <sub>Kr</sub> only	Not reported	Yes	Carmeliet (1992)
Fluoxetine	+	+	Equal	$I_{Na} \leftrightarrow$	Not reported	Yes	Rajamani <i>et al.</i> (2006); Wible <i>et al.</i> (2005)
Ketoconazole	+	+	Equal	$I_{Na} \leftrightarrow$	Not reported	Yes	Dumaine <i>et al.</i> (1998); Mok <i>et al.</i> (2005); Wible <i>et al.</i> (2005); Takemasa <i>et al.</i> (2007)
Pentamidine	_c	+	Trafficking	$I_{kur} \leftrightarrow I_{Ks} \leftrightarrow I_{to} \leftrightarrow $	Kv1.5 ↔	Yes	Cordes et al. (2005) Kuryshev et al. (2005) Wible et al. (2005)
Probucol	_	+	Trafficking	$I_{Na} \leftrightarrow I_{to} \leftrightarrow I_{K1} \leftrightarrow I$		Yes	Guo et al. (2007)
Thioridazine	++	+	< trafficking	'KI ` '		Yes	Drolet <i>et al</i> . (1999); Wible <i>et al</i> . (2005)

Abbreviation: APD, action potential duration; FACS, fluorescence-activated cell sorting;  $I_{Kr}$ , rapid delayed rectifier current carried by hERG; QT, depolarizing–repolarizing time interval on ECG.

<sup>&</sup>lt;sup>a</sup>Effect of prolonged drug treatment on ion-current densities disregarding acute block.

<sup>&</sup>lt;sup>b</sup>Trafficking as detected by biochemical or FACS-based methods.

<sup>&</sup>lt;sup>c</sup>Acute block is observed at concentrations far above clinically relevant exposures only.

which is not obvious for drugs affecting multiple repolarizing and depolarizing ion currents simultaneously. Eventually, drugs may enter the clinical stage without being tested thoroughly for effects on ion channel trafficking.

To illustrate this risk, the characteristics of several drugs that have short-term and long-term effects on  $I_{Kr}$  are shown in Table 1 (see Wible et al. (2005) for a comprehensive screen of 100 compounds). Many drugs display both 'block of function' and a 'reduced density' effect, albeit the latter effect often occurs at lower drug concentrations. The difference in dose dependence further highlights the importance of evaluating the long-term risk for trafficking defects. Some drugs, such as pentamidine, display no direct  $I_{\rm Kr}$  block, but clearly disrupt hERG trafficking at clinically relevant concentrations, which is accompanied by a prolonged APD (Cordes et al., 2005; Kuryshev et al., 2005) and increased QT and pro-arrhythmia (Eisenhauer et al., 1994; Girgis et al., 1997). Furthermore, other ion channels involved in cardiac AP formation are not affected, although trafficking defects for these channels have not been investigated thoroughly. Pentamidine has a reputation of curing critically ill patients with Pneumocystis carinii pneumonia, but making them highly vulnerable for lethal cardiac arrhythmias at the time they leave the intensive care unit (Quadrel et al., 1992), exactly the risk one would predict based on the 'reduced density' hypothesis. From this example, it seems mandatory for the pharmaceutical industry to start screening their compounds for altered ion channel trafficking too.

Finally, as attractive as the simplicity of ectopic expression systems like HEK cells might appear, life is more complex. Screening should take into account other ion channels that function in concert with  $I_{Kr}$ . Proof from cellular APD and animal models should be provided to validate that the observed in vitro effects are truly active and relevant when integrated in the cardiomyocyte AP. As stated by Takemasa et al. (2008), plasma protein binding of ketoconazole is nearly 99%, resulting in effective ketoconozole concentrations in vivo that most likely do not result in either 'block of function' or 'decreased density' effects with respect to hERGmediated  $I_{Kr}$ . Second, there are several examples of drugs displaying blockade of many individual ion currents, but which in concert do not harm the cardiac AP or even relieve some forms of pro-arrhythmia (Verduyn et al., 1995; Oros et al., 2007). Moreover, some drugs like arsenic trioxide block some and activate other repolarizing currents (Drolet et al., 2004) further illustrating the necessity for drug screening in multifactor in vitro test systems and in vivo animal models. The use of pro-arrhythmic models (Thomsen et al., 2006) is advocated because only then a link can be established between drug-induced 'reduced density' and the actual occurrence of torsades de pointes.

## References

Anderson CL, Delisle BP, Anson BD, Kilby JA, Will ML, Tester DJ *et al.* (2006). Most LQT2 mutations reduce Kv11.1 (hERG) current by a class 2 (trafficking-deficient) mechanism. *Circulation* 113: 365–373.

- Carmeliet E (1992). Voltage- and time-dependent block of the delayed K<sup>+</sup> current in cardiac myocytes by dofetilide. *J Pharmacol Exp Ther* **262**: 809–817.
- Cordes JS, Sun Z, Lloyd DB, Bradley JA, Opsahl AC, Tengowski MW *et al.* (2005). Pentamidine reduces *hERG* expression to prolong the QT interval. *Br J Pharmacol* **145**: 15–23.
- Drolet B, Vincent F, Rail J, Chanine M, Deschenes D, Nadeau S *et al.* (1999). Thioridazine lengthens repolarization of cardiac ventricular myocytes by blocking the delayed rectifier potassium current. *J Pharmacol Exp Ther* **288**: 1261–1268.
- Drolet B, Simard C, Roden DM (2004). Unusual effects of a QT prolonging drug, arsenic trioxide, on cardiac potassium currents. *Circulation* **109**: 26–29.
- Dumaine R, Roy M-L, Brown AM (1998). Blockade of HERG and Kv1.5 by ketoconazole. *J Pharmacol Exp Ther* **286**: 727–735.
- Eckhardt LL, Rajamani S, January CT (2005). Protein trafficking abnormalities: a new mechanism in drug-induced long QT syndrome. *Br J Pharmacol* **145**: 3–4.
- Eisenhauer MD, Eliasson AH, Taylor AJ, Coyne Jr PE, Wortham DC (1994). Incidence of cardiac arrhythmias during intravenous pentamidine therapy in HIV-infected patients. *Chest* **105**: 389–394.
- Ficker E, Kuryshev YA, Dennis AT, Obejero-Paz C, Wang L, Hawryluk P *et al.* (2004). Mechanisms of arsenic-induced prolongation of cardiac repolarization. *Mol Pharmacol* **66**: 33–44.
- Fitzgerald PT, Ackerman MJ (2005). Drug-induced torsades de pointes: The evolving role of pharmacogenetics. *Heart Rhythm* 2: \$30–\$37.
- Girgis I, Gualberti J, Langan L, Malek S, Mustaciuolo V, Costantino T et al. (1997). A prospective study of the effect of IV pentamidine therapy on ventricular arrhythmias and QTc prolongation in HIVinfected patients. Chest 112: 646–653.
- Guo J, Massaeli H, Li W, Xu J, Luo T, Shaw J *et al.* (2007). Identification of  $I_{\rm Kr}$  and its trafficking disruption induced by probucol in cultured neonatal rat cardiomyocytes. *J Pharmacol Exp Ther* **321**: 911–920.
- Hancox JC, Mitcheson JS (2006). Combined hERG channel inhibition and disruption of trafficking in drug-induced long QT syndrome by fluoxetine: a case-study in cardiac safety pharmacology. Br J Pharmacol 149: 457–459.
- Kuryshev YA, Ficker E, Wang L, Hawryluk P, Dennis AT, Wible BA et al. (2005). Pentamidine-induced long QT syndrome and block of hERG trafficking. J Pharmacol Exp Ther 312: 316–323.
- Mok N-S, Lo Y-K, Tsui P-T, Lam C-W (2005). Ketoconazole induced Torsades de Pointes without concomitant use of QT intervalprologing drug. *J Cardiovasc Electrophysiol* **16**: 1375–1377.
- Oros A, Antoons G, Oosterhoff P, Houtman MJC, Attevelt NJM, Beekman JDM *et al.* (2007). The capacity of the heart to withstand an arrhythmogenic challenge increases with flunarizine which can be quantified using beat-to-beat variability of repolarization. *Eur Heart J* 28: 400–401.
- Quadrel MA, Atkin SH, Jaker MA (1992). Delayed cardiotoxicity during treatment with intravenous pentamidine: two case reports and a review of the literature. *Am Heart J* **123**: 1377–1379.
- Rajamani S, Eckhardt LL, Valdivia CR, Klemens CA, Gillman BM, Anderson CL *et al.* (2006). Drug-induced long QT syndrome: hERG K<sup>+</sup> channel block and disruption of protein trafficking by fluoxetine and norfluoxetine. *Br J Pharmacol* **149**: 481–489.
- Ridley JM, Milnes JT, Duncan RS, McPate MJ, James AF, Witchel HJ *et al.* (2006). Inhibition of the HERG K<sup>+</sup> channel by the antifungal drug ketoconazole depends on channel gating and involves the S6 residue F656. *FEBS Lett* **580**: 1999–2005.
- Roden DM, Balser JR, George Jr AL, Anderson ME (2002). Cardiac ion channels. *Annu Rev Physiol* **64**: 431–475.
- Stansfeld PJ, Gedeck P, Gosling M, Cox B, Mitcheson JS, Sutcliffe MJ (2007). Drug block of the hERG potassium channel: insight from modeling. *Proteins* **68**: 568–580.
- Steele DF, Eldstrom J, Fedida D (2007). Mechanisms of cardiac potassium channel trafficking. *J Physiol* **582**: 17–26.
- Sun H, Liu X, Xiong Q, Shikano S, Li M (2006). Chronic inhibition of cardiac Kir2.1 and hERG potassium channels by celastrol with dual effects on both ion conductivity and protein trafficking. *J Biol Chem* 281: 5877–5884.

- Takemasa H, Nagatomo T, Abe H, Kawakami K, Igarashi T, Tsurugi T *et al.* (2008). Coexistence of hERG current block and disruption of protein trafficking in ketoconazole-induced long QT syndrome. *Br J Pharmacol* **153**: 439–447 (this issue).
- Thomsen MB, Matz J, Volders PGA, Vos MA (2006). Assessing the proarrhythmic potential of drugs: current status of models and surrogate parameters of torsades de pointes arrhythmias. *Pharmacol Ther* 112: 150–170.
- Verduyn SC, Vos MA, Gorgels AP, Van der Zande J, Leunissen JD, Wellens HJ (1995). The effect of flunarizine and ryanodine on
- acquired torsades de pointes arrhythmias in the intact canine heart. *J Cardiovasc Electrophysiol* 6: 189–200.
- Walker VE, Atanasiu R, Lam H, Shrier A (2007). Co-chaperone FKBP38 promotes HERG trafficking. *J Biol Chem* **282**: 23509–23516.
- Wang L, Wible BA, Wan X, Ficker E (2007). Cardiac glycosides as novel inhibitors of human *Ether-a-go-go*-related gene channel trafficking. *J Pharmacol Exp Ther* **320**: 525–534.
- Wible BA, Hawryluk P, Ficker E, Kuryshev YA, Kirch G, Brown AM (2005). HERG-Lite: a novel comprehensive high-throughput screen for drug-induced hERG risk. *J Pharmacol Toxicol Methods* **52**: 136–145.